

GENOMICA

Genómica: Subdisciplina de la Genética que tiene que ver con la clonación y caracterización molecular de genomas completos.

- **Genómica estructural:** Es la caracterización de la naturaleza física de genomas completos (secuenciación, ensamblaje, anotación)
- **Genómica funcional:** Es la caracterización de transcriptomas y proteomas. Conjunto de genes de un genoma que determinan proteínas y de los patrones globales de expresión génica.

Avances claves en genómica

año

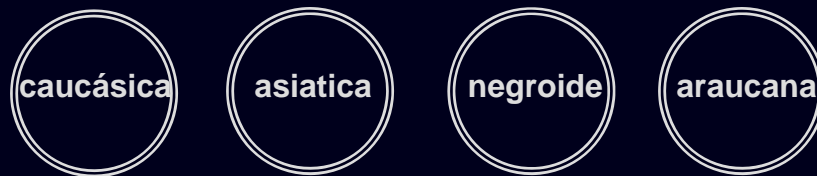
- • Double helix structure of DNA - Watson and Crick 1953
- • Sintesis in vitro de RNA – Severo Ochoa 1959
- • Descubrimiento del código genético (M.Nirenberg y H. J. Matthaei) 1968
- • DNA sequencing - Sanger method 1973
- • Detection of specific DNA sequences - Southern 1975
- • DNA amplification by PCR - Mullis 1988
- • Array-based genetic determinations – P.Brown 1995
- • Sequencing of the human genome- J.Ventner et al. 2001

Análisis Genómico

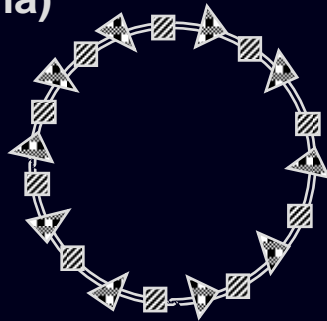
- Conocer los códigos contenidos en el genoma en un organismo y establecer su relación con otros genomas. (genómica comparada, filogenia, etc)



- Establecer la variabilidad de cada genoma (polimorfismo, SNPs)



- Identificar regiones codificadoras y reguladoras (anotación del genoma)

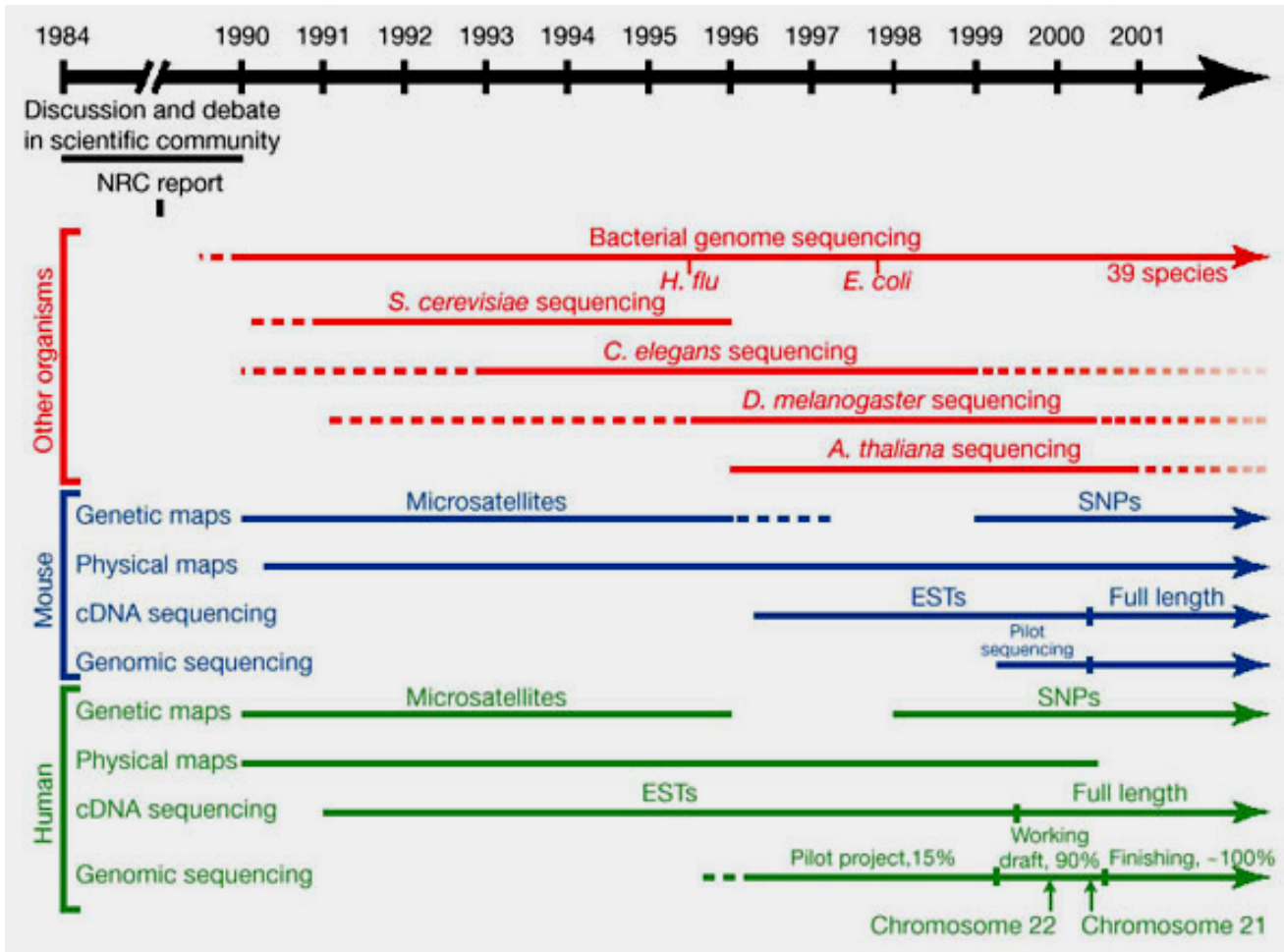


- ▨ Codificadora, aquella que es transcrita en RNA
- Reguladora, aquella que regula la transcripción

QUE SE ESPERA DE LA GENOMICA FUNCIONAL

Organism	# genes	% of genes with inferred function	Completion date of genome
<i>E. coli</i>	4288	60	1997
yeast	6,600	40	1996
<i>C. elegans</i>	19,000	40	1998
<i>Drosophila</i>	12-14K	25	1999
<i>Arabidopsis</i>	25,000	40	2000
mouse	~30,000?	10-20	2003
human	30-40,000	10-20	2002

HISTORIA DE LA GENOMICA



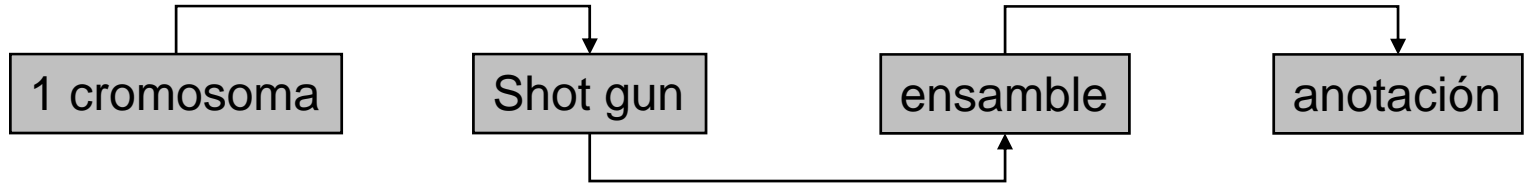
27-10-2008
766-NCBI

Completo
July 2007 (mm9)
assembly 2,700x10⁶

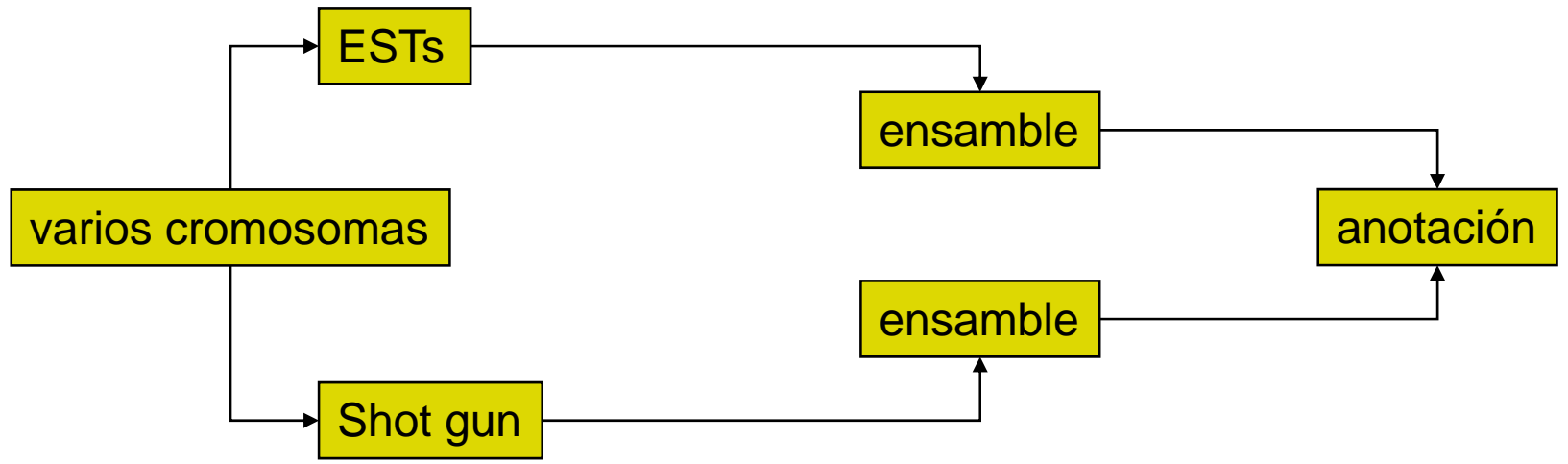
Finalizado
NCBI Build 36.1, Mar.
2006 Assembly (hg18)
3,080x10⁶

Otros 14 mamíferos; 10 vertebrados, 13 insectos; entre otros eucariontes han sido secuenciado total o parcialmente

Bacterias



Eucariontes



Aislar el material genético de un organismo

1,5-4,5 x10⁶ bp

Trozar el DNA (Shot-gun)

1-5 Kb y 40-60 Kb

Cloning

genoteca

1.000-100.000 clones

Secuenciación automática

10000 corridas (ambos extremos)

Lectura de secuencia nucleotídica

600-800 pb corridas

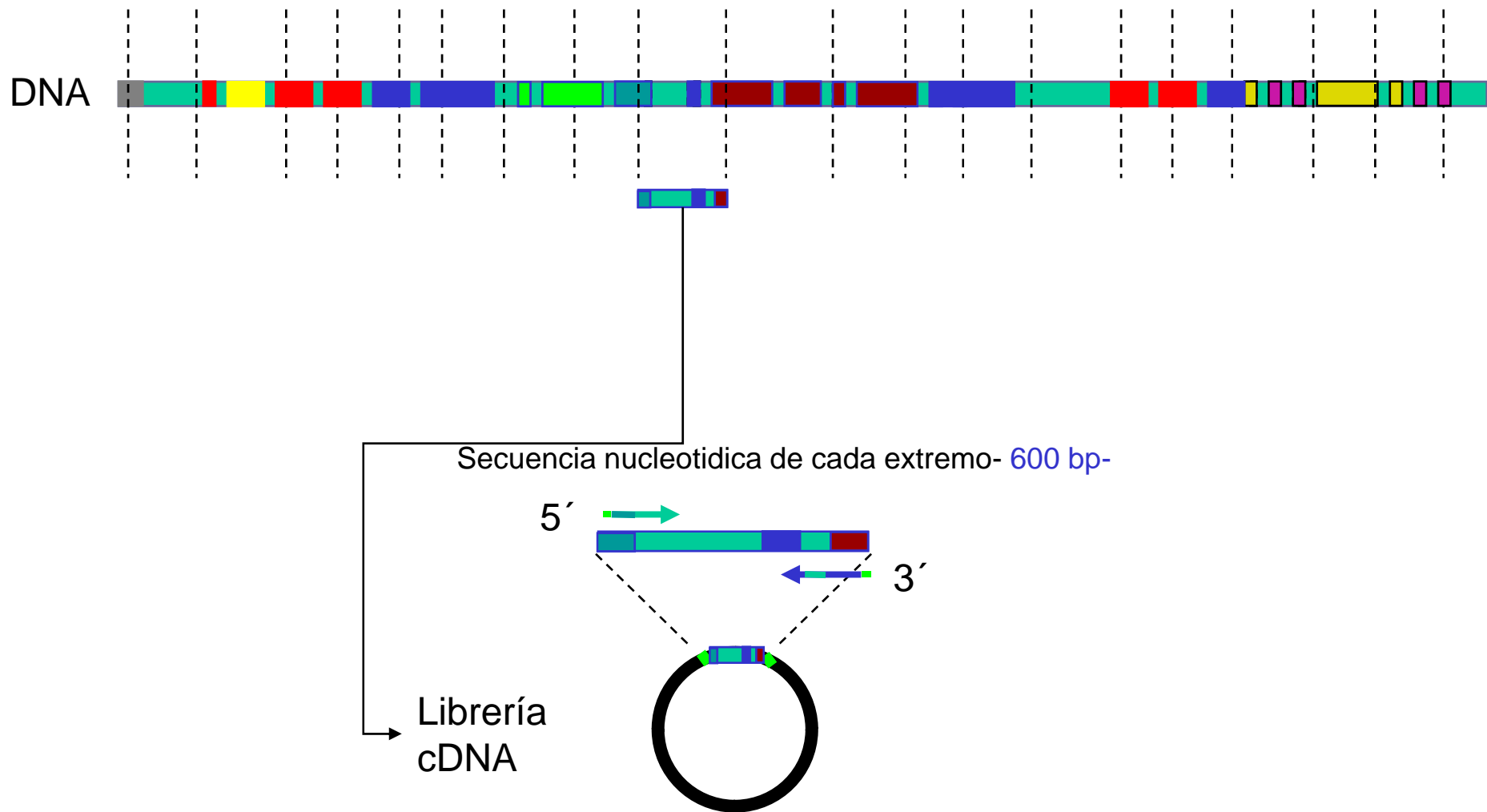
12 x10⁶ bp

ensamblaje

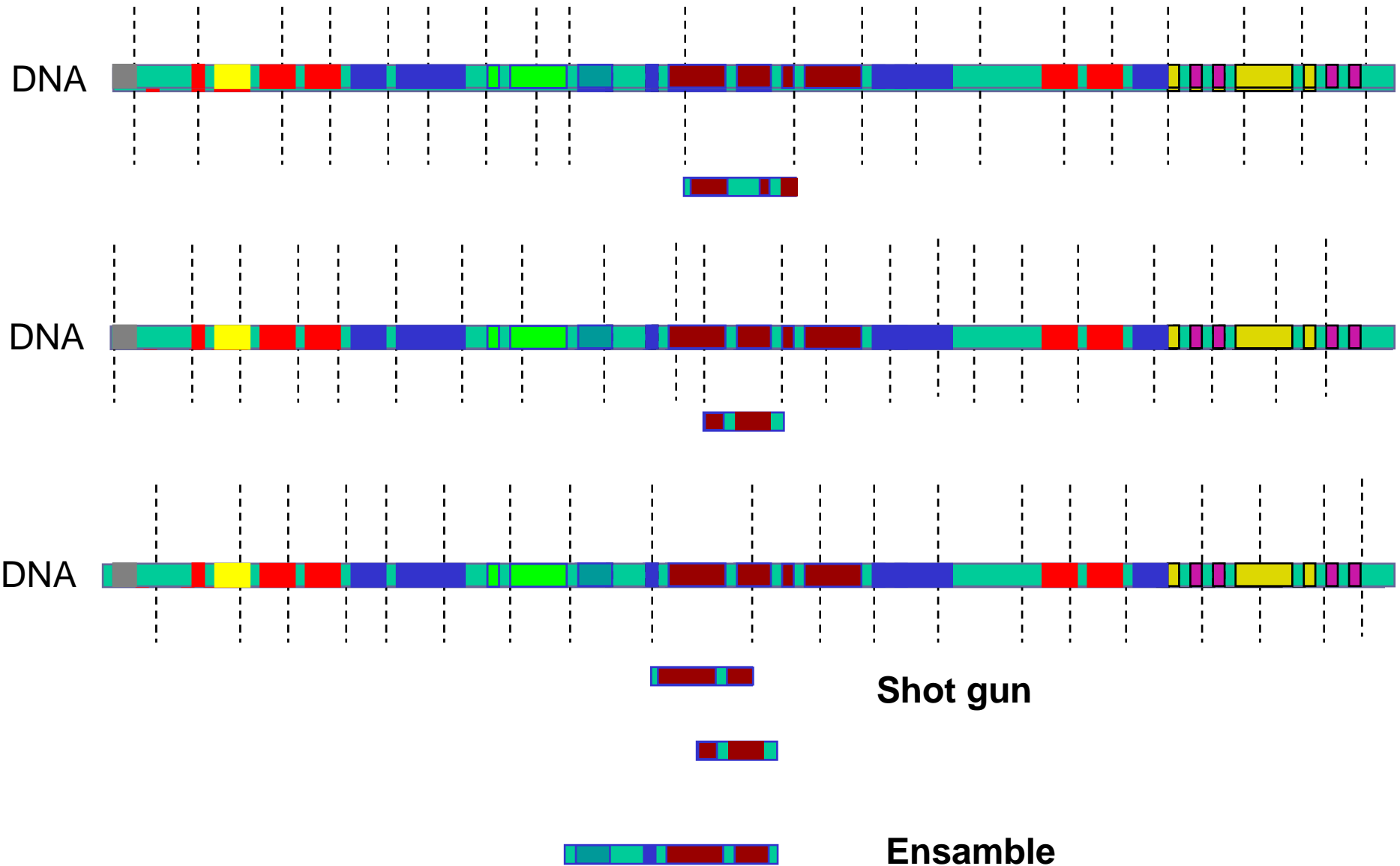
8-3 veces

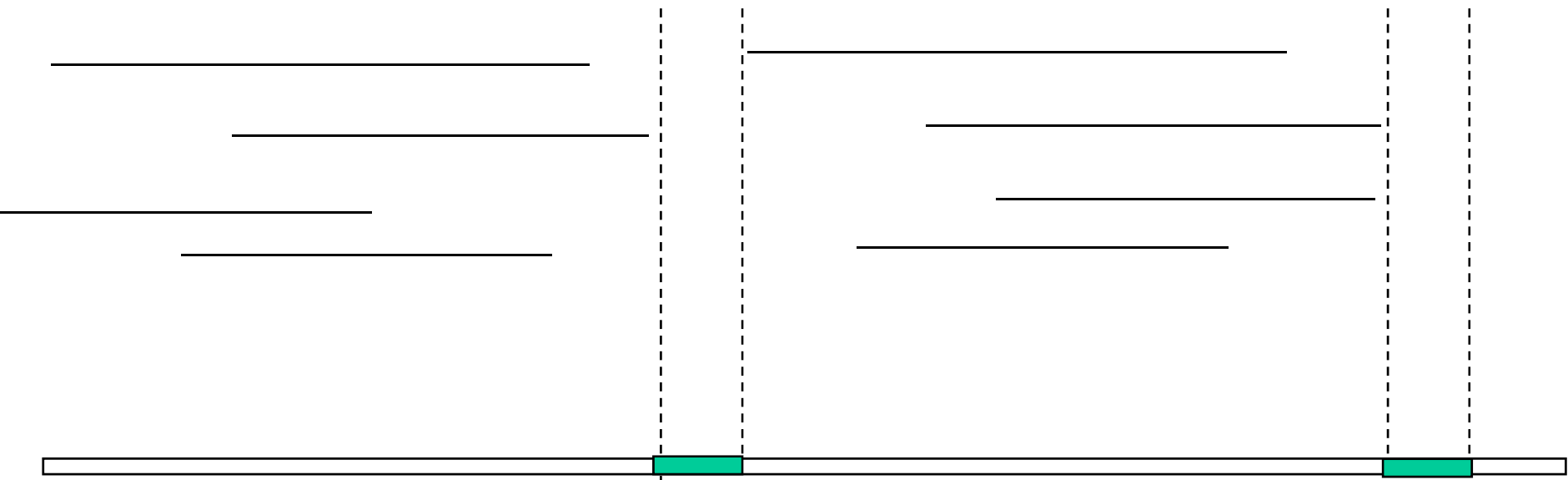
Shot gun

Cortes con enzimas de restricción

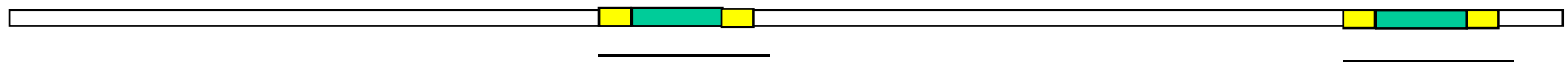


Shot gun



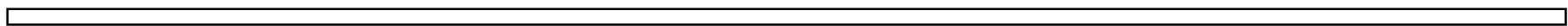


Recuperación de fragmentos vía PCR



1

2

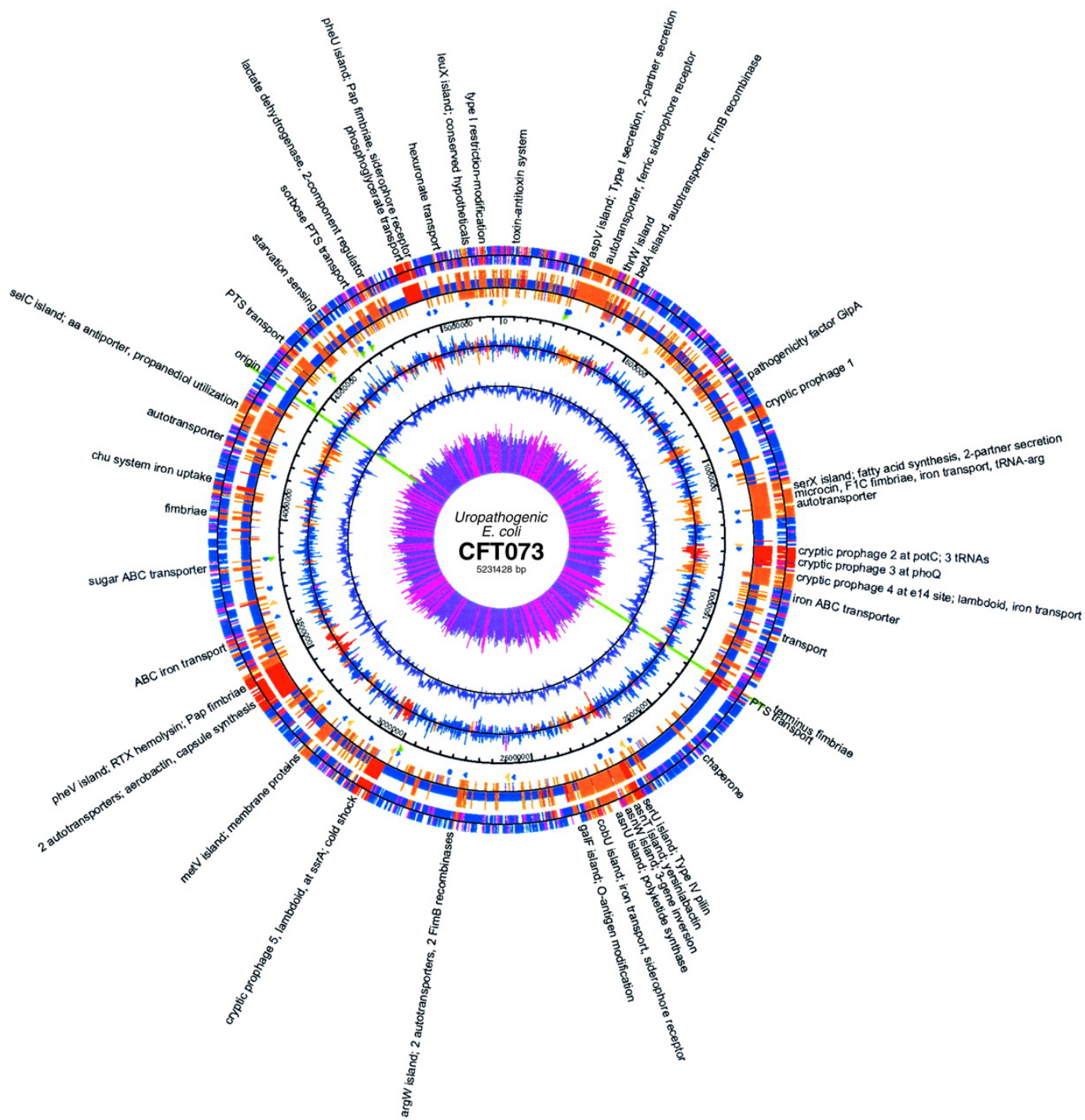


Repetición del protocolo de secuenciación con nuevos clones.

Table 1. General Features of the *Thermoanaerobacter tengcongensis* Genome

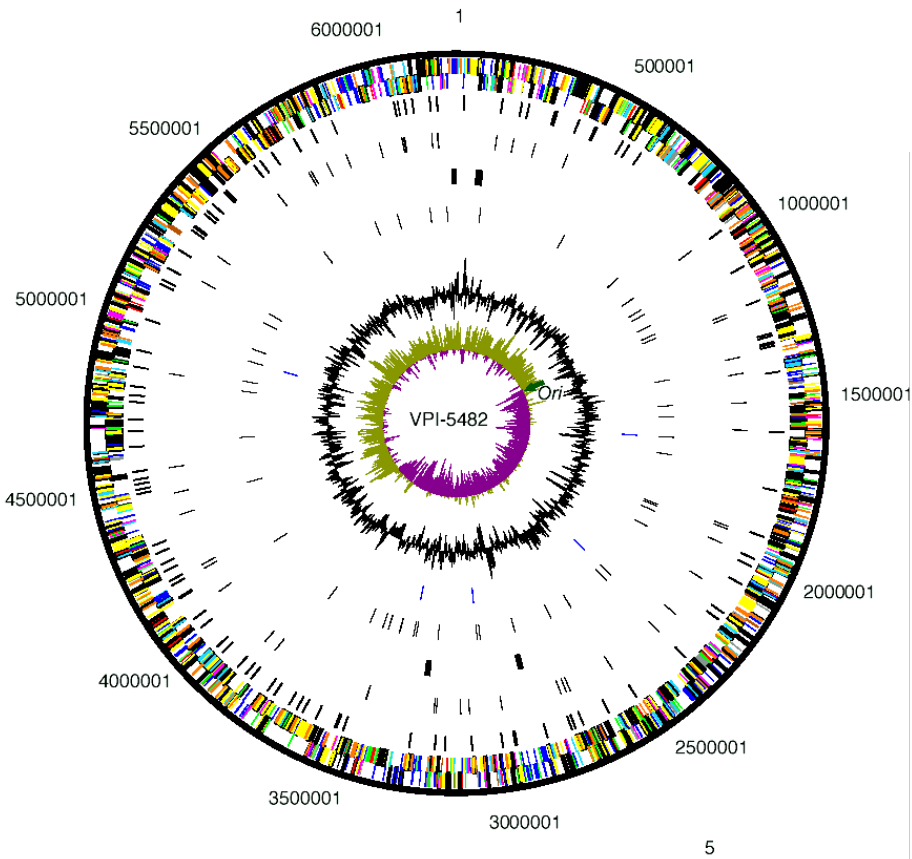
Genome size (bp)		2,689,445
G+C content		37.6%
Protein coding		87.1%
Average CDS length (bp)		905
Predicted CDS		2,588
	Known proteins	1494 (57.8%)
Homologous to	Protein domains/motifs	270 (10.4%)
	Hypothetical proteins	301 (11.6%)
No homology		523 (20.2%)
Stable RNAs		0.9%
rRNA operons		4
rRNAs		55
Major repetitive elements		
Short noncoding repeats		0.5%
Long coding repeats		8.6%

CDS, coding sequences.

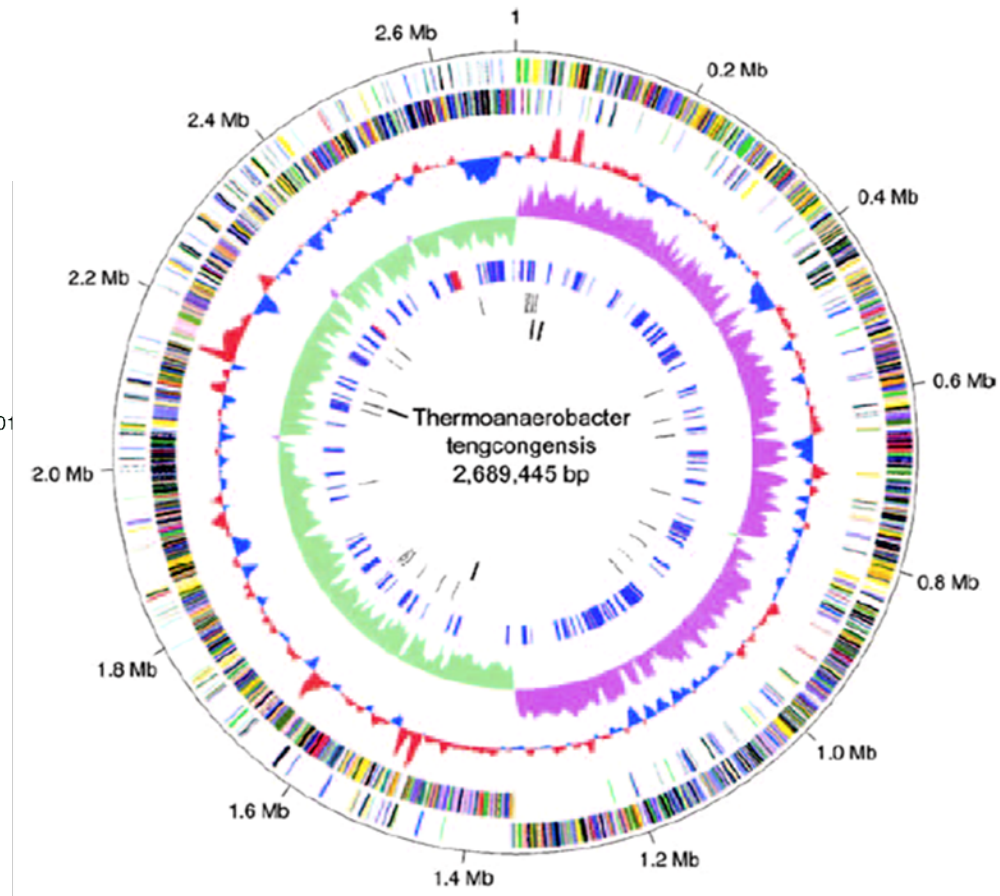


Map of the CFT073 genome and comparison with K-12 strain MG1655. The outer circle shows ORFs, colored according to the K-12 comparison in the second circle, where DNA regions are shown: blue, backbone, i.e., E. coli near match to MG1655; red, CFT073 islands (insertions); orange, islands (substitutions replacing K-12 segments); violet, K-12 islands. ORFs in the outer ring that span island-backbone junctions are pink. Third circle, RNAs: green, rRNA operons; blue, tRNAs; gold, miscellaneous RNAs. Fourth circle, scale in bp. Fifth circle, GC skew calculated for each ORF >100 aa, colored according to the same scheme of the ORF circle and plotted around the mean. Sixth circle, GC skew calculated over the whole sequence (window, 10 kb) plotted around the mean. Seventh circle, codon-adaptation index CAI (inverse, 1-CAI is plotted); pink rays indicate CAI values <0.2; purple rays, values >0.2. The pink rays can be seen to correspond with islands. A detailed linear map with ORF annotations is available at www.genome.wisc.edu. Maps were created by GENVISION from DNASTAR.

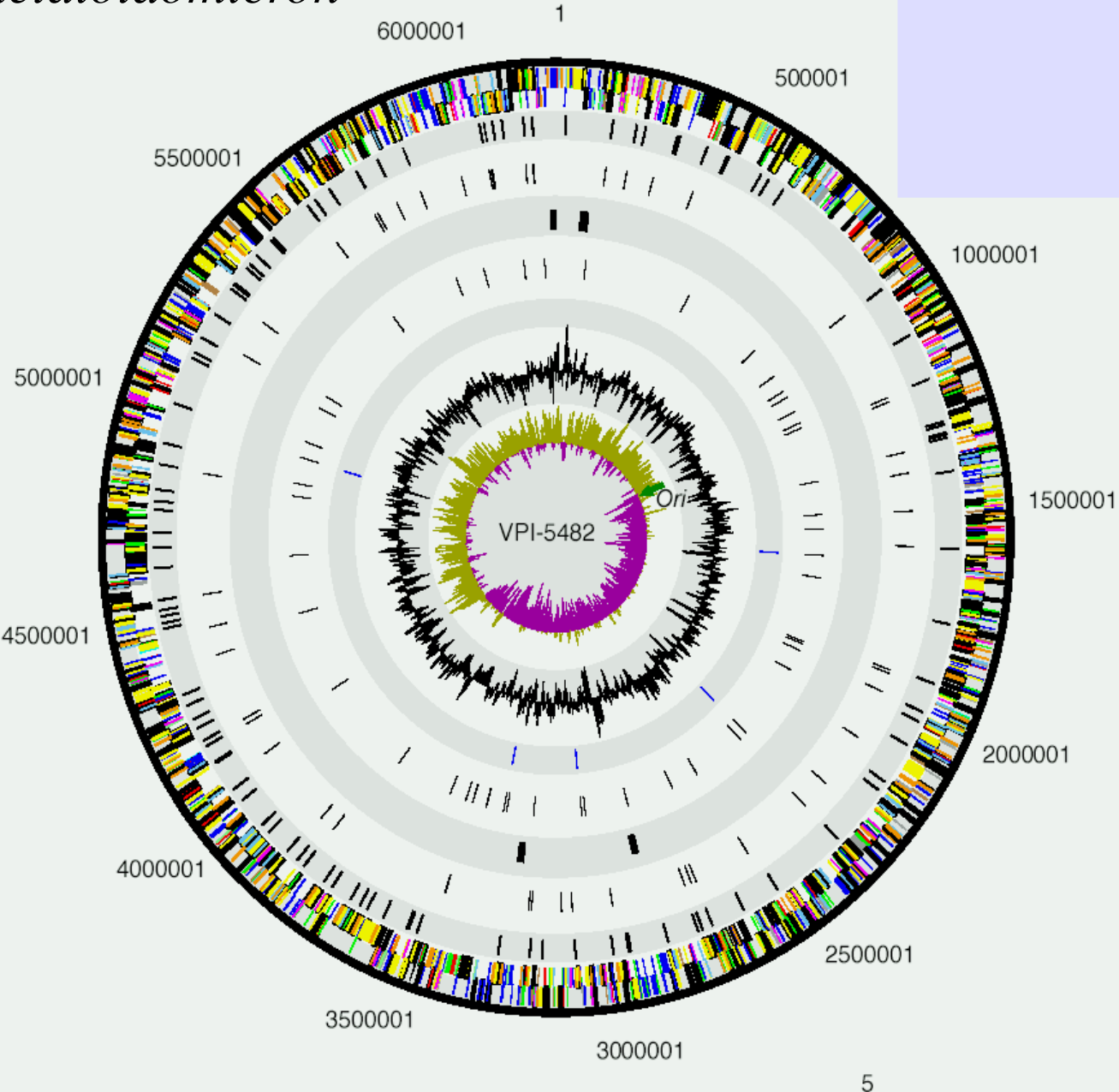
B. thetaiotaomicron



T. tengcongensis



B. thetaiotaomicron



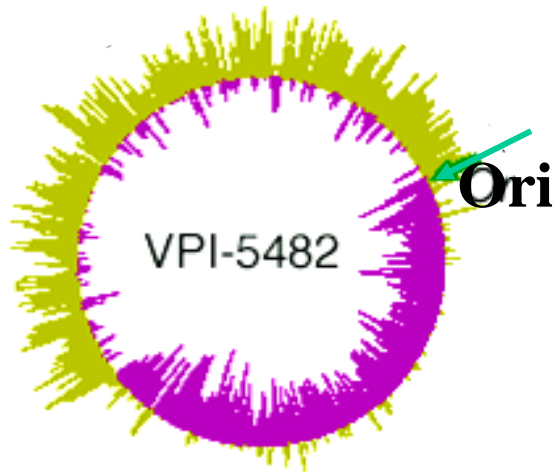
Secuencia lineal de un cromosoma bacteriano

B. thetaiotaomicron

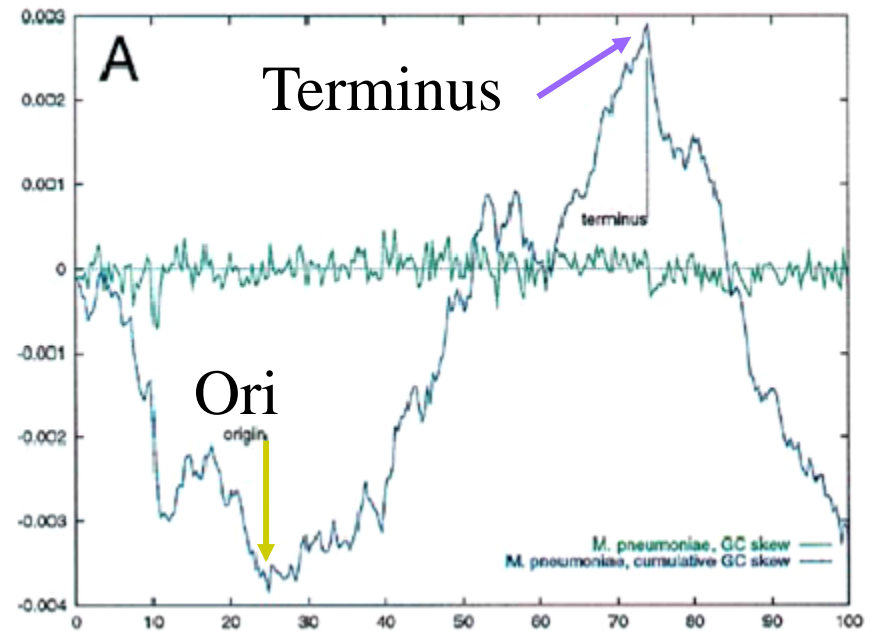
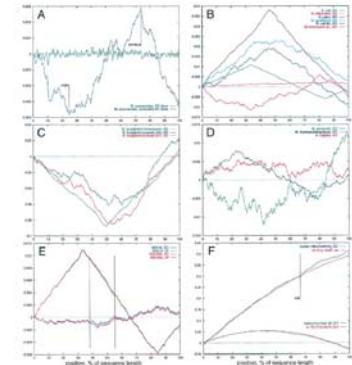
5'

3'

Origin of DNA replication:



GC skew.



1. GC nucleotide skew $[(G-C)/(G+C)]$ analysis.

- 2 Identification by similarity, in this case we evaluated the co-localisation of several genes found around the origin of others bacterias. Typically bacteria contain the genes ***dnaA***, ***dnaN***, ***recF*** and ***gyrB***, however the *Bacillus subtilis* chromosome include only the genes *dnaA* and *dnaN* while ***rpmH*** and ***recF*** are not include (Kuroda et al., 2001. The Lancet, 357: 1225), in *E. coli* *dnaN* and *gyrB* are present, *dnaA* is located 600bp away from *dnaN* – *gyrB*, and *recF* gene is missing. (Zawilaketal., 2001. Nucleid Acids Research **29**:2251)

- 3 In addition, we need evaluate if the replication origins region, contain short AT rich sequences and named **DnaA box** characteristic of bacterial (in *E. coli* and in *H. Pylori* there are 5 DnaA box).

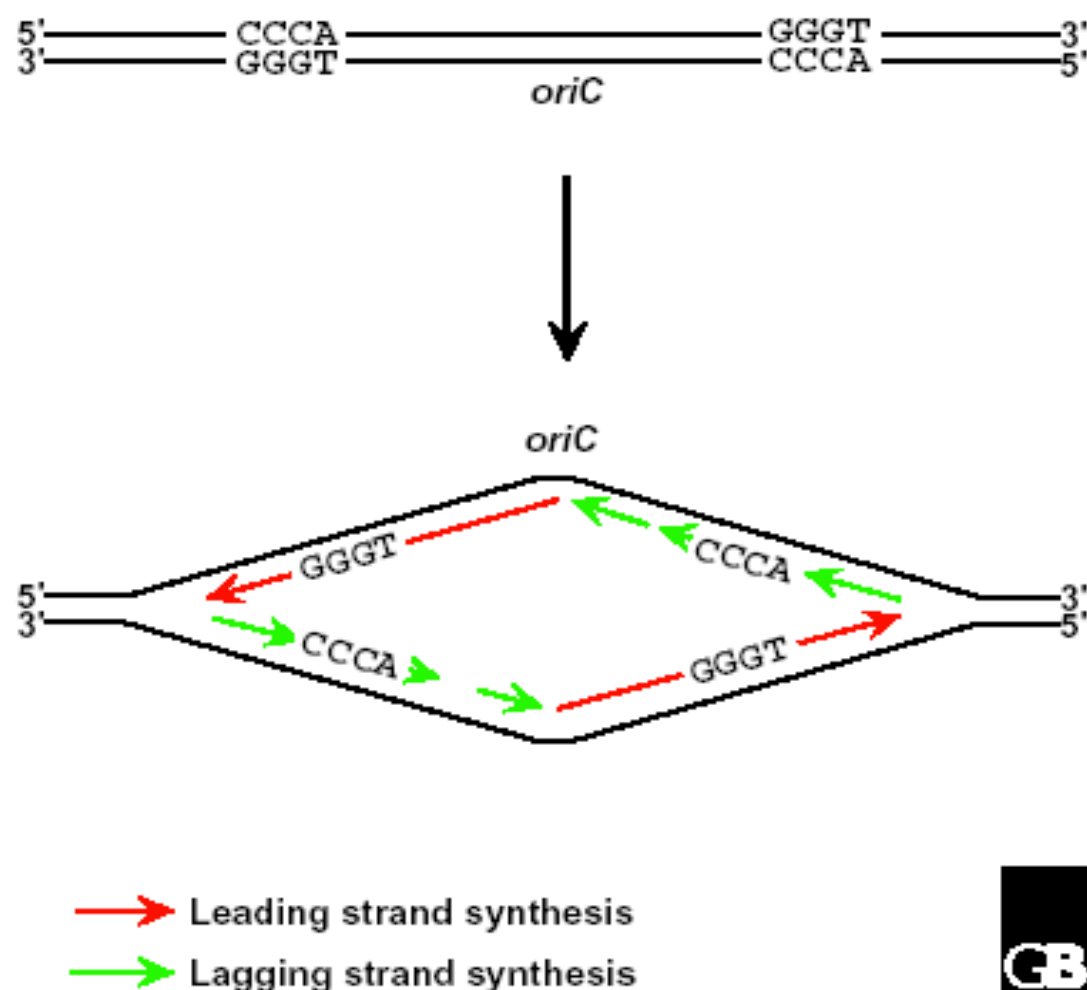
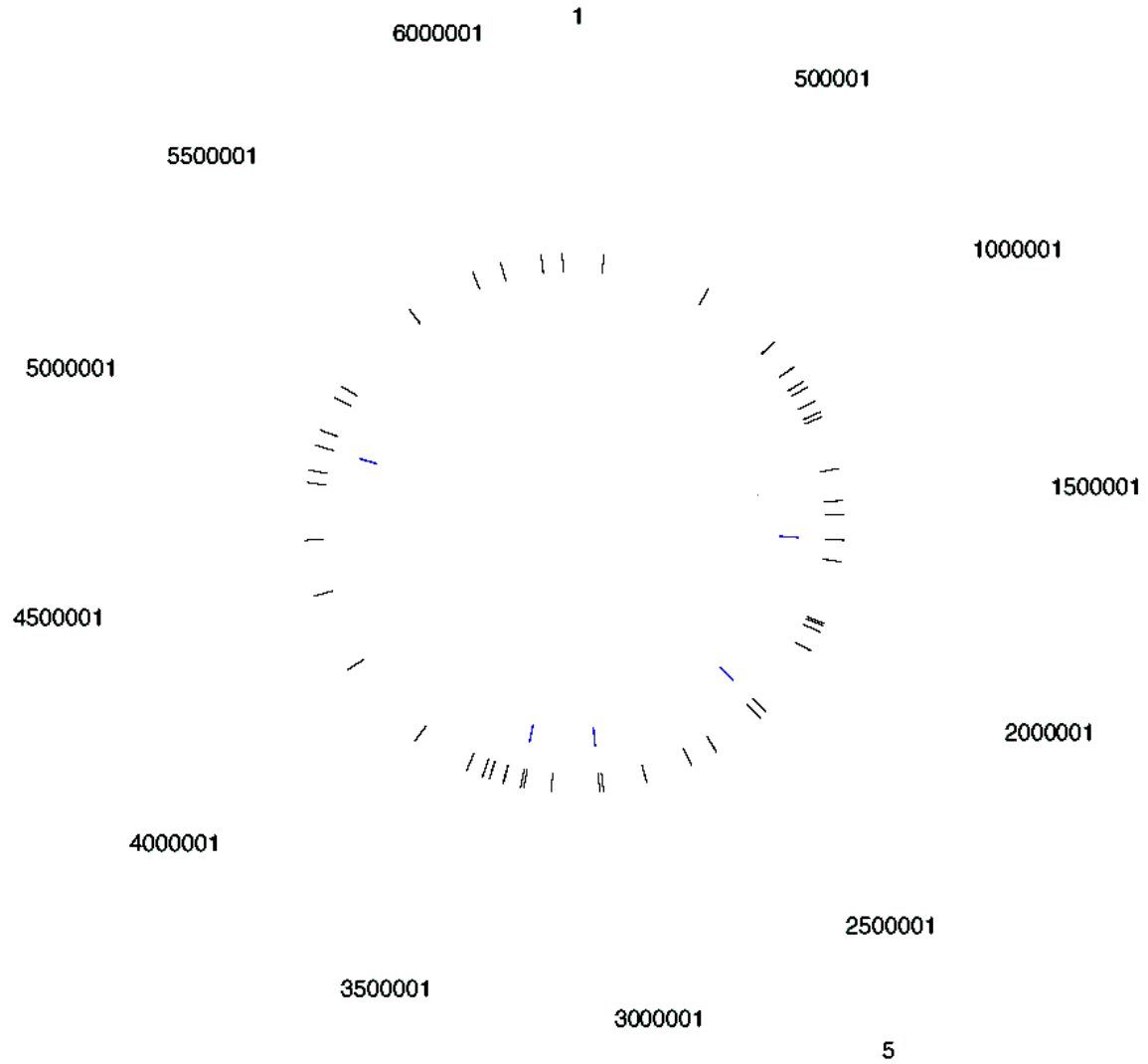


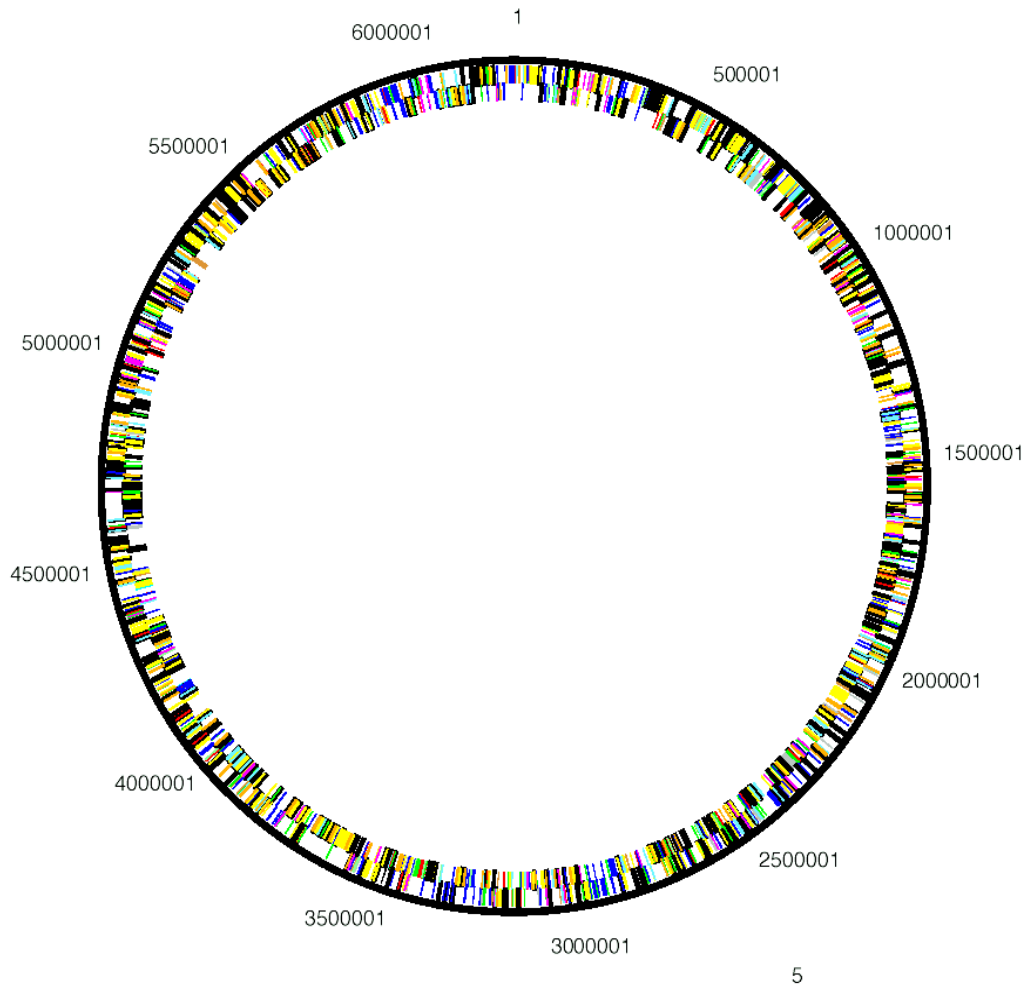
Figure 2

A bi-directional replication fork: DNA replicated by lagging-strand synthesis on one side of the origin will be replicated by leading-strand synthesis on the other side. In bacteria, there is a switch in the strand bias of guanine content at the origin. Myllykallio *et al.* [1] measured the strand bias resulting from GGGT as illustrated.

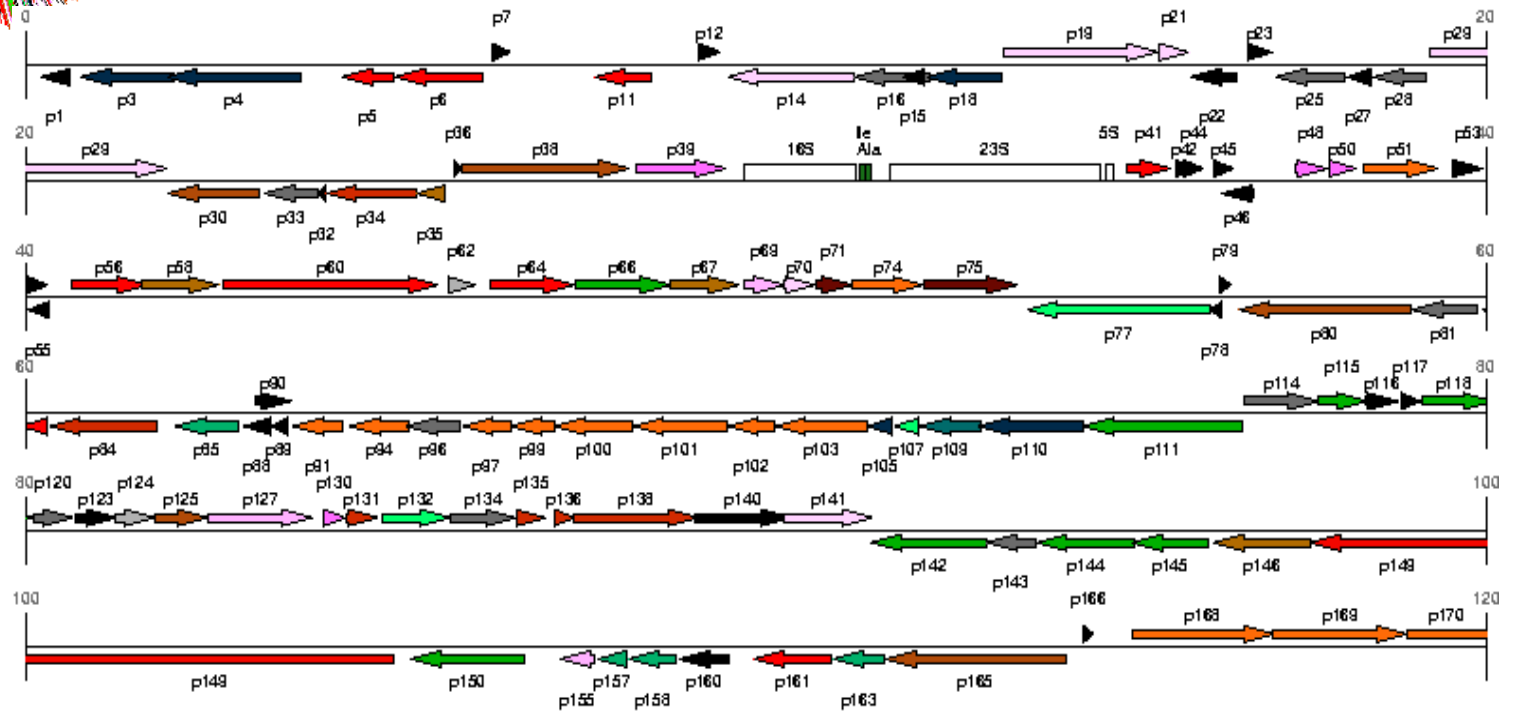
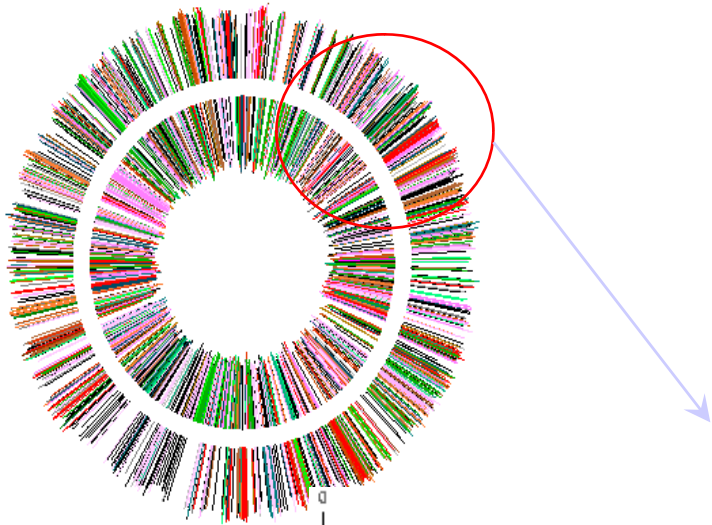
rRNA: BLASTN

tRNA genes: tRNA-Scan



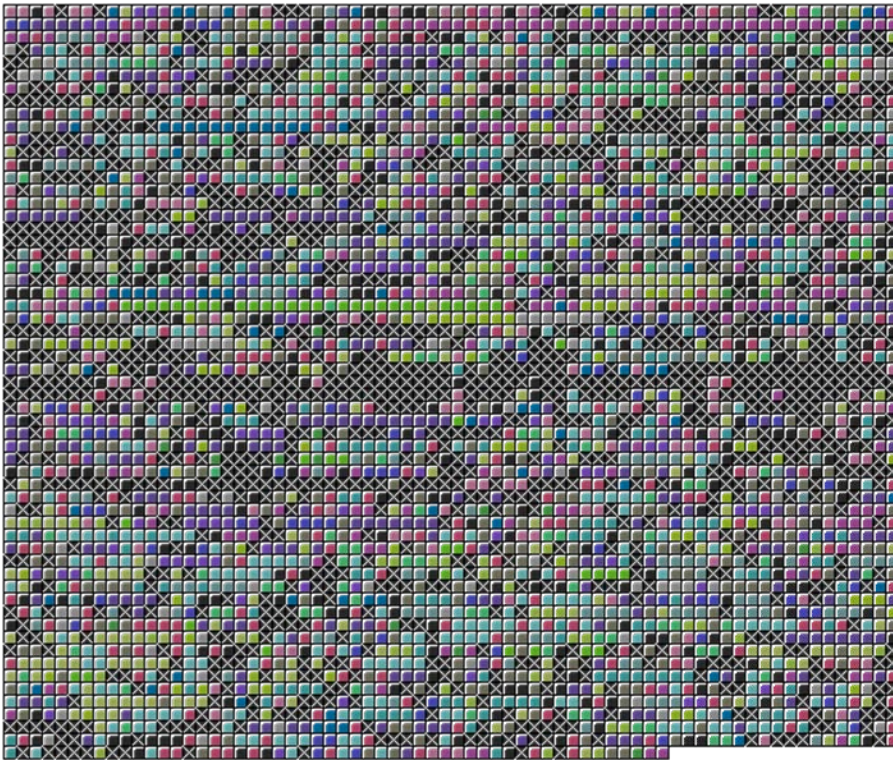


1. Identificación de marcos de lectura por hebra. (utilizando **GLIMMER**, ORPHEUS and CRITICA .
2. Verificación por blast. % recuperado x homolg.
3. Revisión manual
4. Genes que codifican para proteínas con <60-100 codones que no presentan homologías con un cierto umbral de *E value* (10^3) deben ser eliminadas.
5. Clasificación de funciones (gen-ontology) por grandes grupos.
6. Mapa físico en megabases

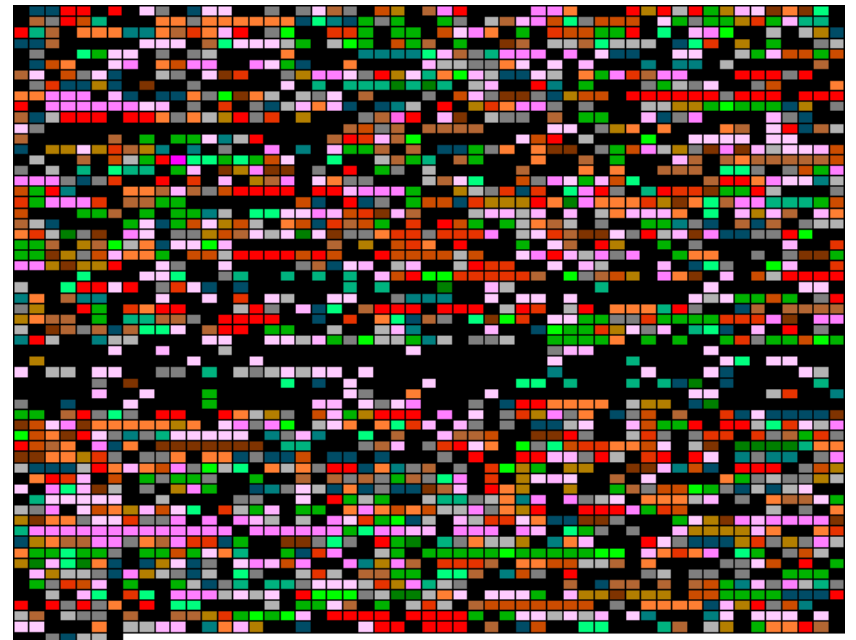


Anotación funcional de genomas de bacterias

Anotación NCBI de *Bacillus subtilis*:



Anotación LBMG bacteria biominera

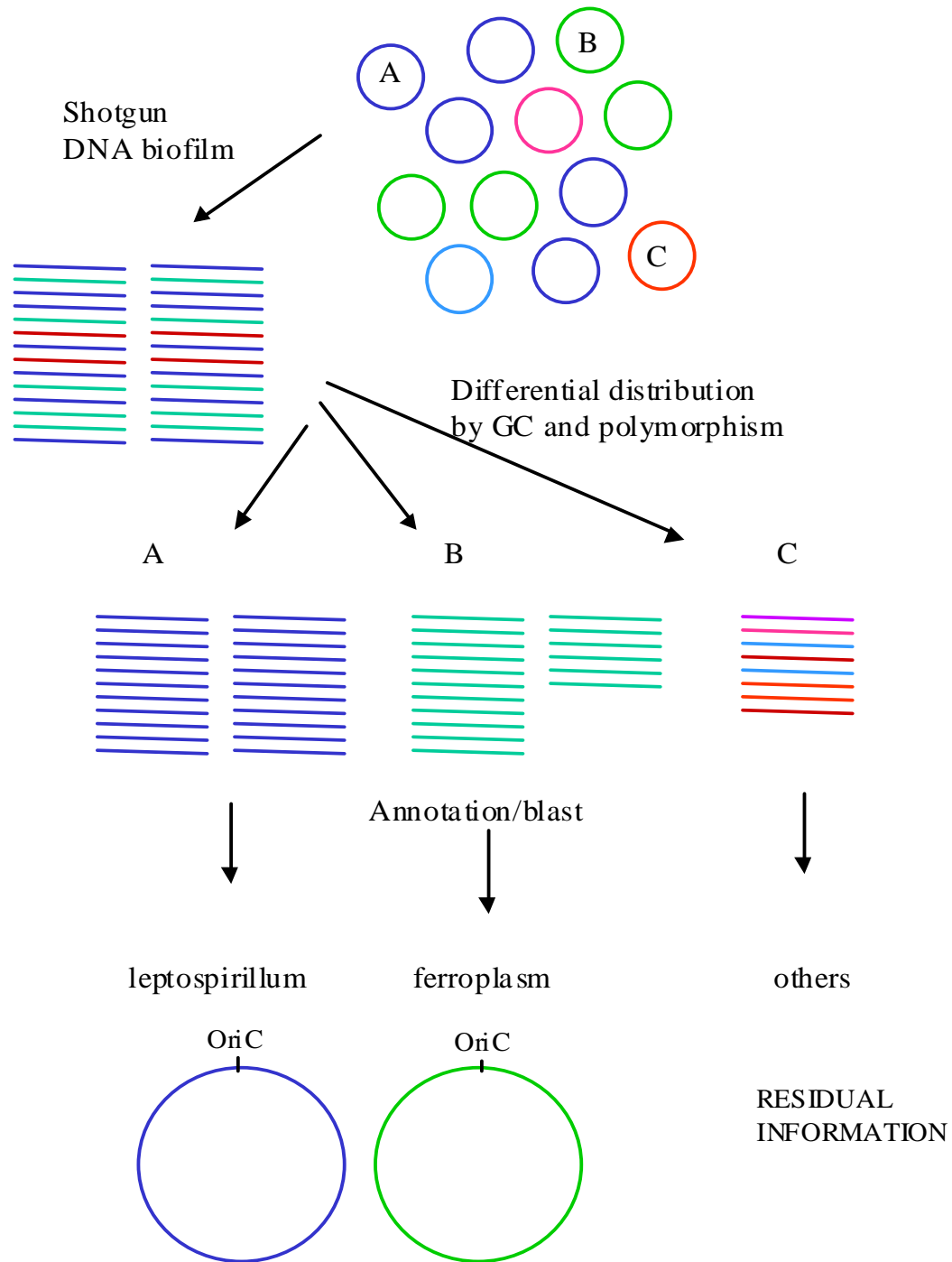


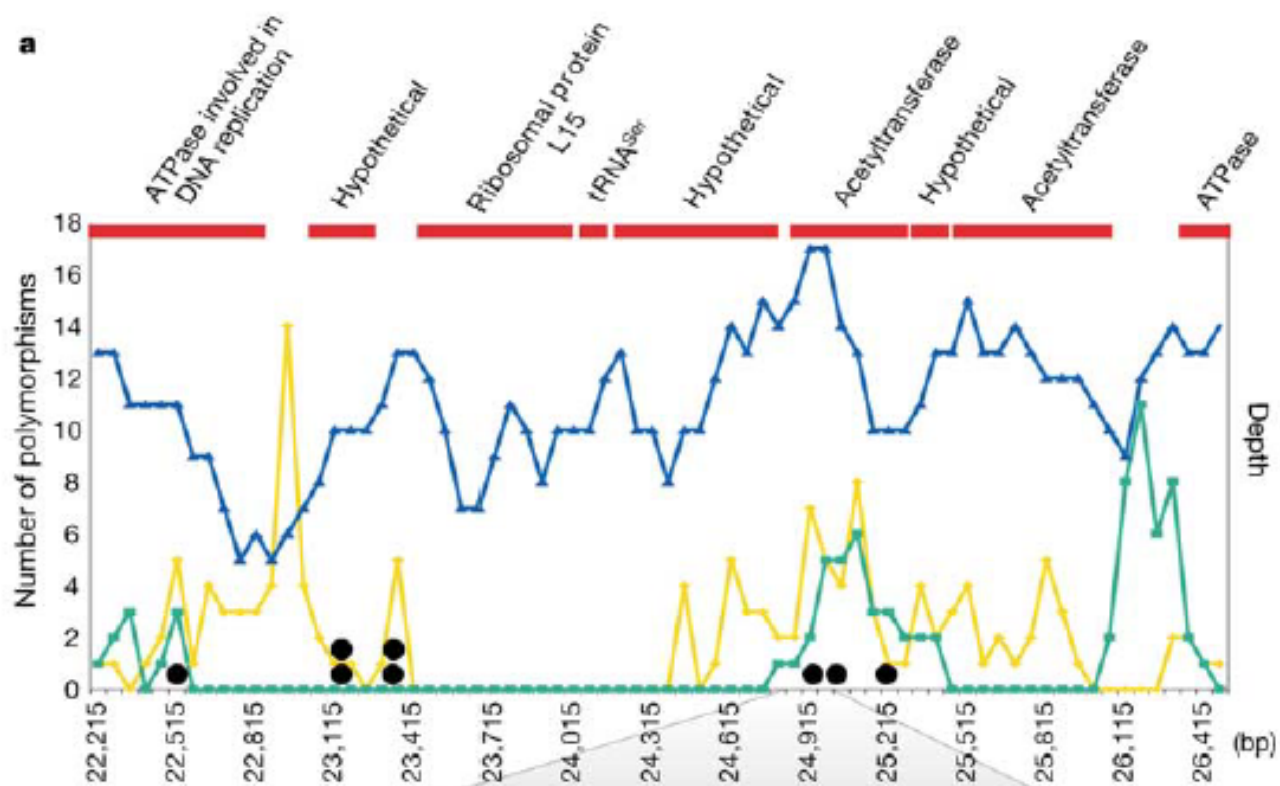
Community structure and metabolism through reconstruction of microbial genomes from the environment

Gene W. Tyson¹, Jarrod Chapman^{3,4}, Phillip Hugenholtz¹, Eric E. Allen¹, Rachna J. Ram¹, Paul M. Richardson⁴, Victor V. Solovyev⁴, Edward M. Rubin⁴, Daniel S. Rokhsar^{3,4} & Jillian F. Banfield^{1,2}

¹*Department of Environmental Science, Policy and Management, ²Department of Earth and Planetary Sciences, and ³Department of Physics, University of California, Berkeley, California 94720, USA*

⁴*Joint Genome Institute, Walnut Creek, California 94598, USA*





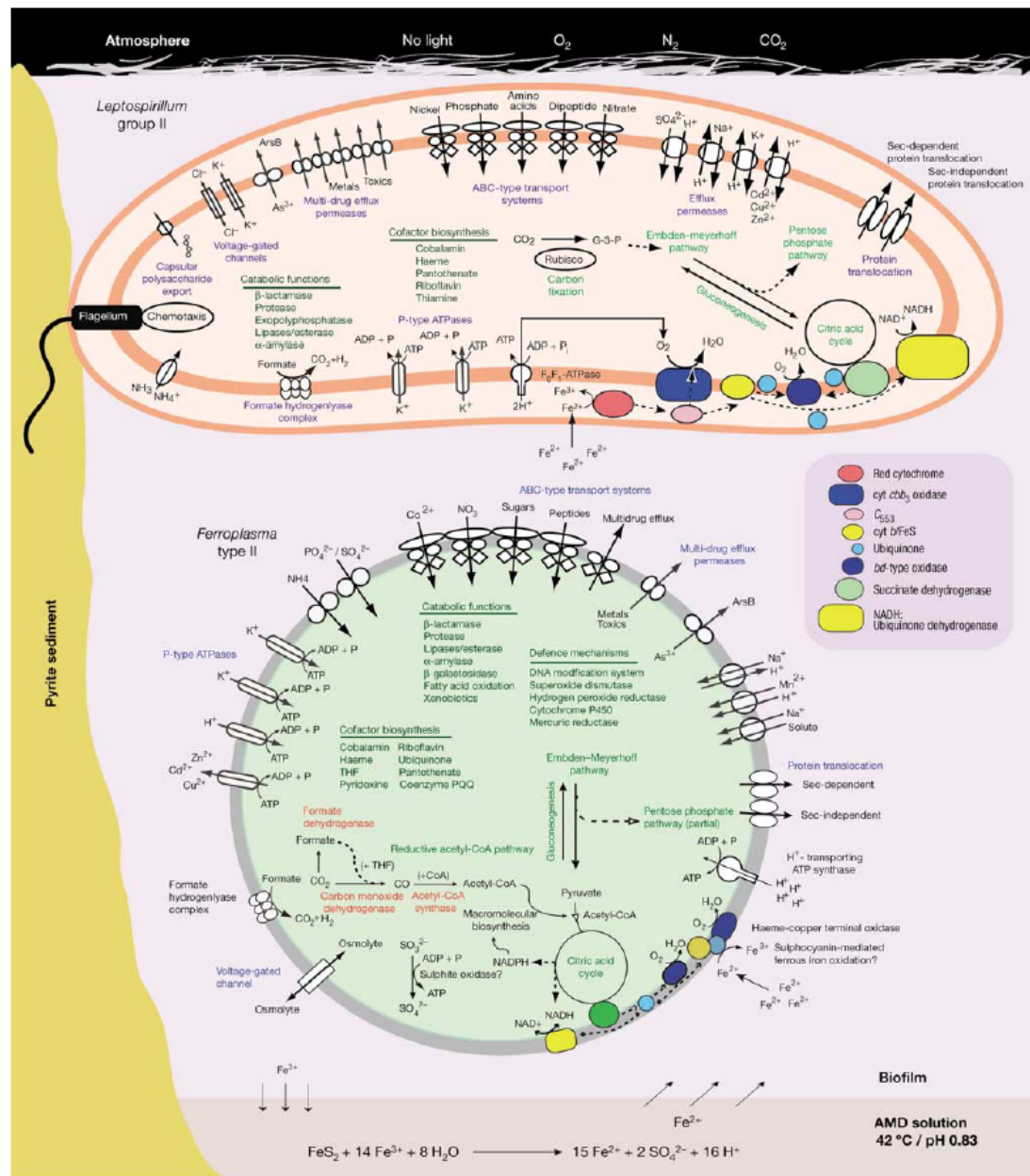
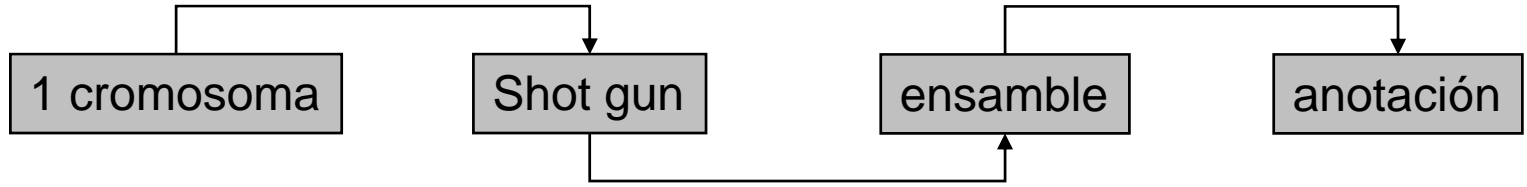


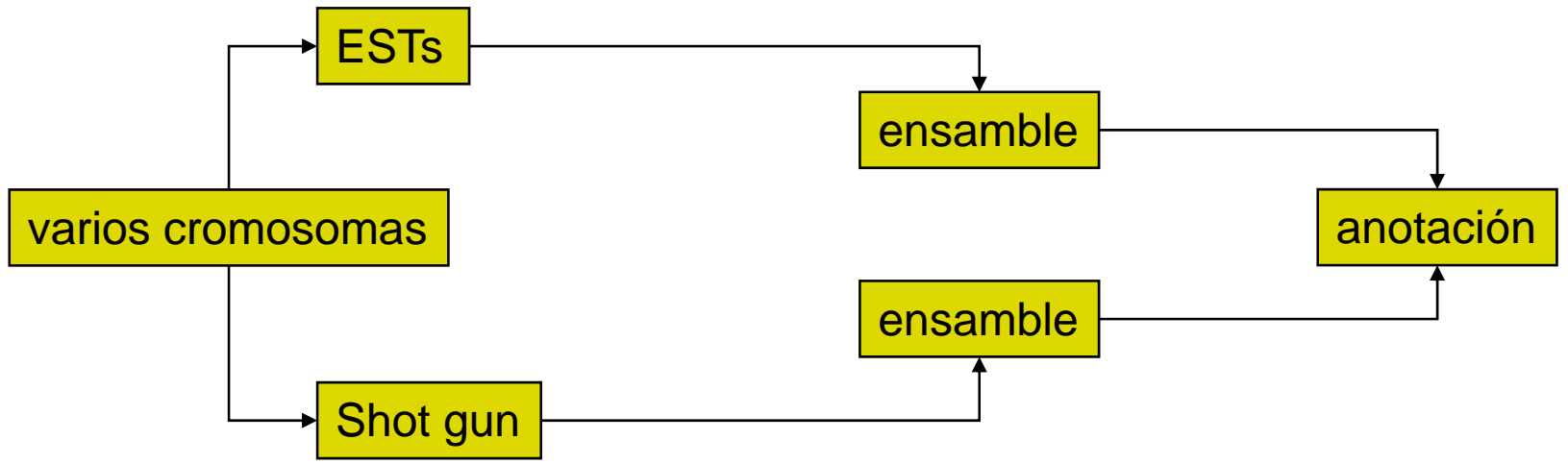
Figure 4 Cell metabolic cartoons constructed from the annotation of 2,180 ORFs identified in the *Leptospirillum* group II genome (63% with putative assigned function) and 1,931 ORFs in the *Ferroplasma* type II genome (58% with assigned function). The cell cartoons are shown within a biofilm that is attached to the surface of an acid mine

drainage stream (viewed in cross-section). Tight coupling between ferrous iron oxidation, pyrite dissolution and acid generation is indicated. Rubisco, ribulose 1,5-bisphosphate carboxylase-oxygenase. THF, tetrahydrofolate.

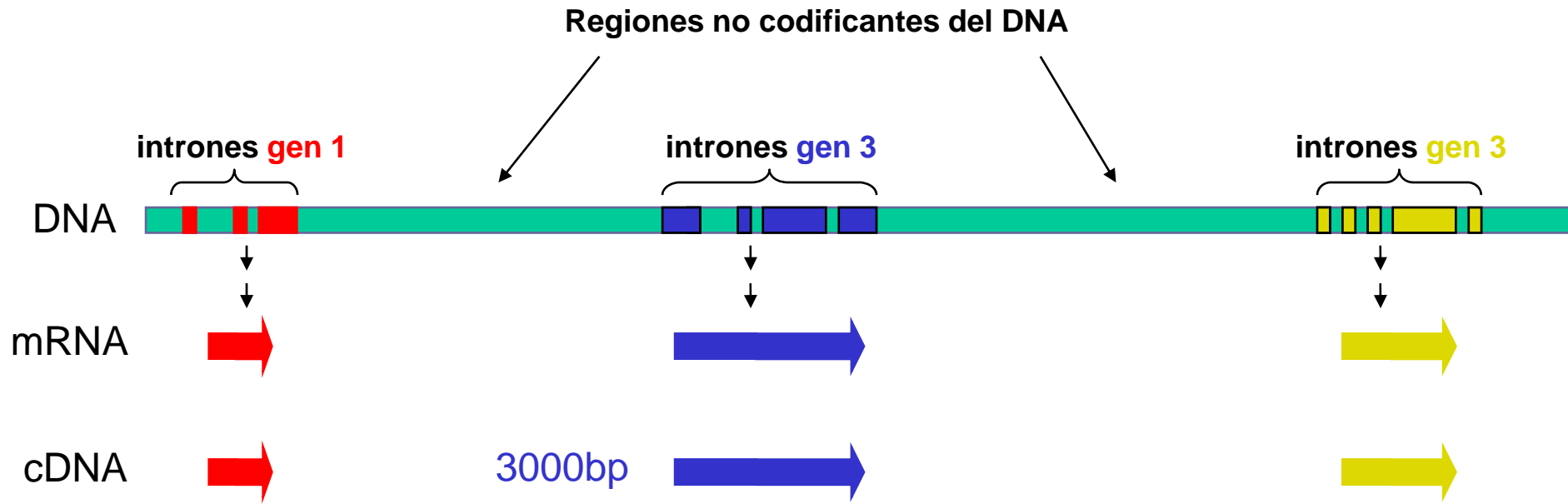
Bacterias



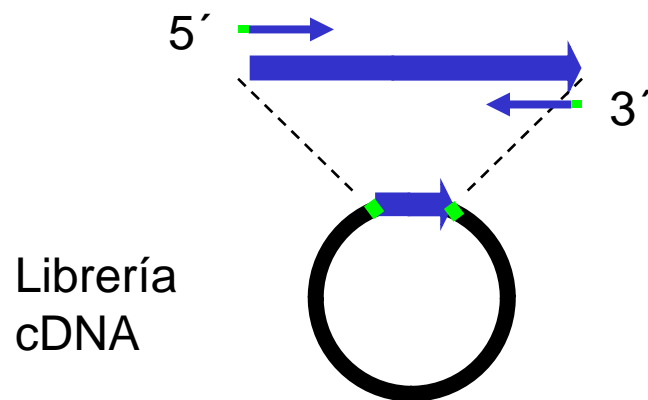
Eucariontes



ESTs

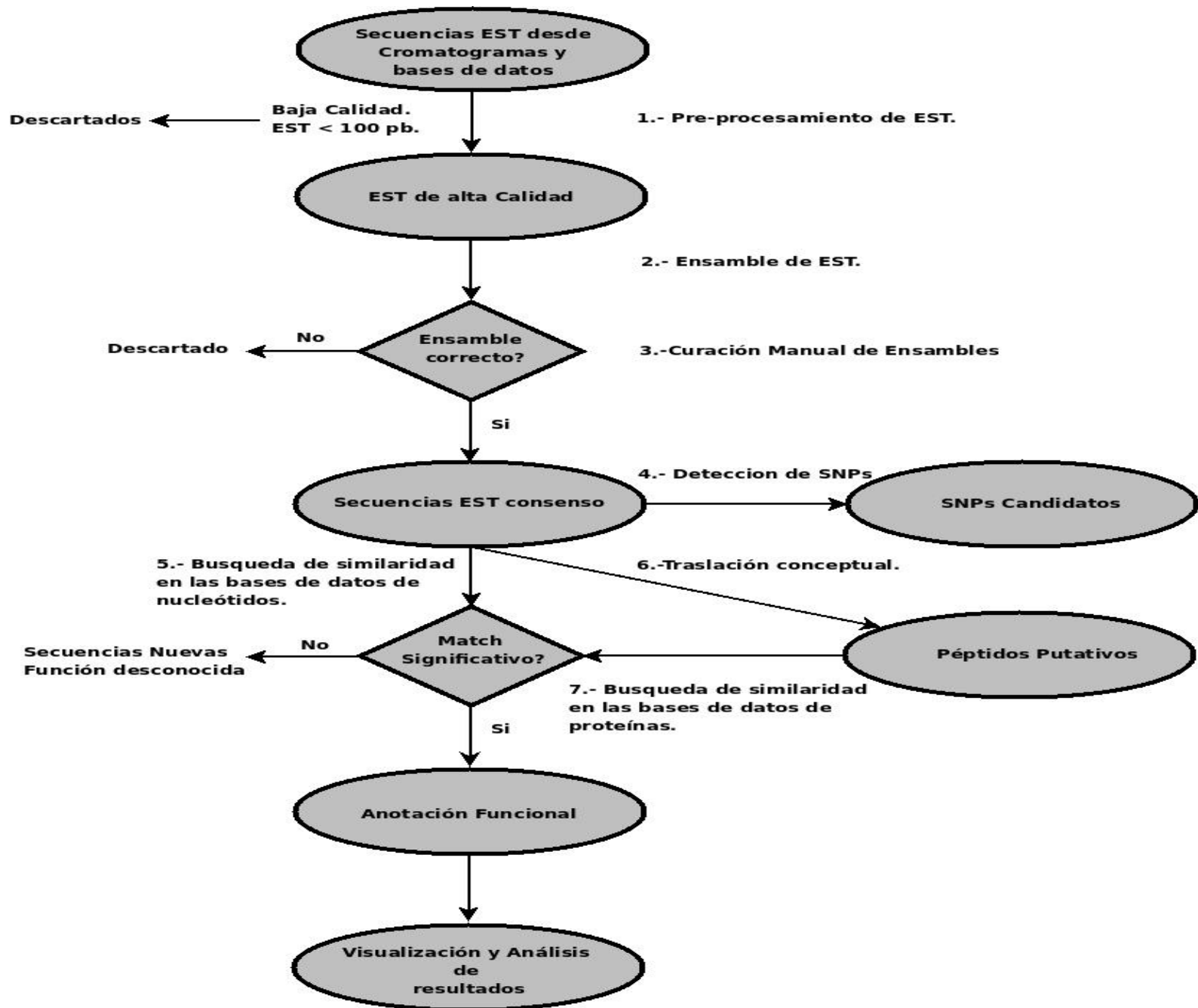


Secuencia nucleotidica de cada extremo- 600 bp-



Avances proyecto genómica de salmones

Nombre : Alex Di Génova.



Bases de Datos

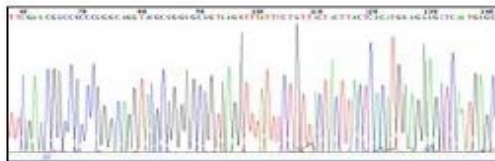
- cGRASP v/s NCBI:
 - Secuencias ESTs Salmo salar:

	Total	Particular	Común
NCBI	433337	139843	293494
cGRASP	487522	194028	293494
TOTAL	293494	333871	627365

Ensamble

- Pre-Ensamble:
 - Determinación de Químicas de los cromatogramas.
 - Enmascarar Vector
 - Utilización de Univec (2862 sec).
 - PblueScriptIIISK+, pCMVsport6 ect.
 - Crossmatch.
 - Enmascarar Poly-A
 - Seqclean y Masked.

Ensamble



```
ACATGGAGGAGCTCAATGAGCTCATTACGACAGAATCCACAACAAGA  
TGATCTTCTTCTCCCTTTTCGTCCCTACTTATCACCAGTAGAAACTGGA  
TAGAAACATTTCCATTATATTACGTTATCAAGCTAGGCATTCTTCTCT  
AAACTGAAGTTCTGGTTCAAGAGGAAACACCAAAAAGCTCC
```

EST desde cromatogramas

EST desde base de datos

```
ACATGGAGGAGCTCAATGAGCTCATTACGACAGAATCCACAACAAGA  
TGATCTTCTTCTCCCTTTTCGTCCCTACTTATCACCAGTAGAAACTGGA  
TAGAAACATTTCCATTATATTACGTTATCAAGCTAGGCATTCTTCTCT  
AAACTGAAGTTCTGGTTCAAGAGGAAACACCAAAAAGCTCC
```

Identificar y remover
la secuencia del vector
(UNIVC y EMVEC)

```
ACATGGAGGAGCTCAATGAGCTCATTACGACAGAATCCACAACAAGA  
TGATCTTCTTCTCCCTTTTCGTCCCTACTTATCACCAGTAGAAACTGGA  
TAGAAACATTTCCATTATATTACGTTATCAAGCTAGGCATTCTTCTXX  
XXXXXXXXXXXXXXXXXXXXXXXXXXXX
```

Enmascaramiento de las
secuencias de baja
complejidad

```
ACATGGAGGAGCTCAATGAGCTCATTACGACAGAATCCACAACAANN  
NNNNNNNNNNNNNNNNCTTTTCGTCCCTACTTATCACCAGTAGAAACTGGA  
TAGAAACATTTCCATTATATTACGTTATCAAGCTAGGCATTCTTCTXX  
XXXXXXXXXXXXXXXXXXXXXXXXXXXX
```

Secuencias de alta calidad para ensamblar

Ensamble

```

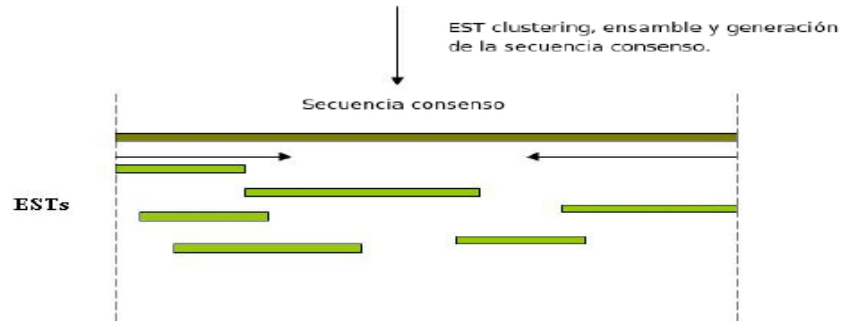
ACATGGAGGAGCTCAATGAGCTCATTACGACAGAATCCACAACANNH
NNNNNNNNNNNNNNCTTTTCGTCCCTACTTATCACCAGTAGAAAACCTGGA → EST 1
TAGAAACATTTCCATTATATTACGTTATCAAGCTAGGCATTCTTCTXX
XXXXXXXXXXXXXXXXXXXXXXXXXXXX
  
```

```

          ACATGGAGGAGCTCAATGAGCTCATTACGACAGAATCCACAACANNH
EST 2 ←  HNNNNNNNNNNNNNNCTTTTCGTCCCTACTTATCACCAGTAGAAAACCTGGA
          TAGAAACATTTCCATTATATTACGTTATCAAGCTAGGCATTCTTCTXX
          XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX
  
```

```

ACATGGAGGAGCTCAATGAGCTCATTACGACAGAATCCACAACANNH
NNNNNNNNNNNNNNCTTTTCGTCCCTACTTATCACCAGTAGAAAACCTGGA → EST 3
TAGAAACATTTCCATTATATTACGTTATCAAGCTAGGCATTCTTCTXX
XXXXXXXXXXXXXXXXXXXXXXXXXXXX
  
```



Un ejemplo del análisis

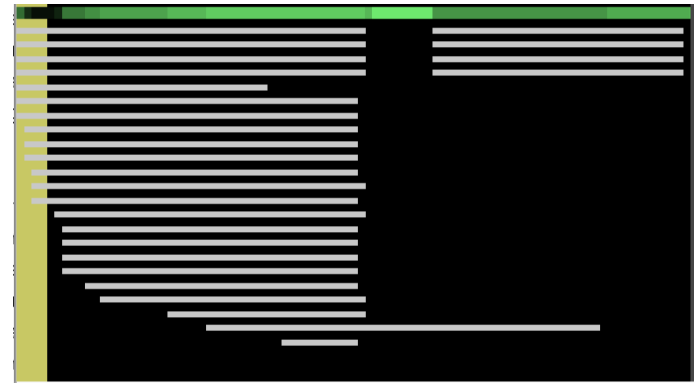
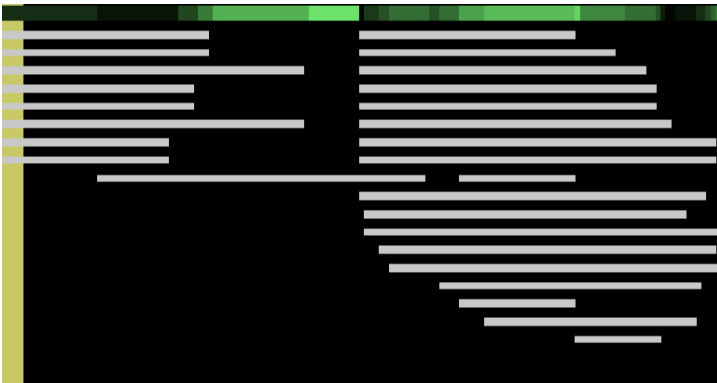
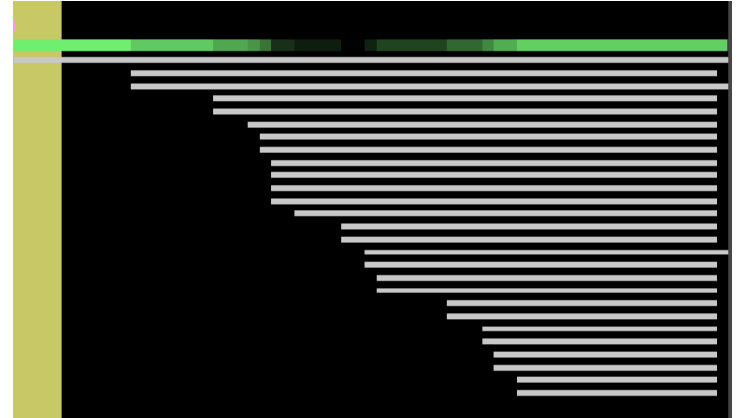
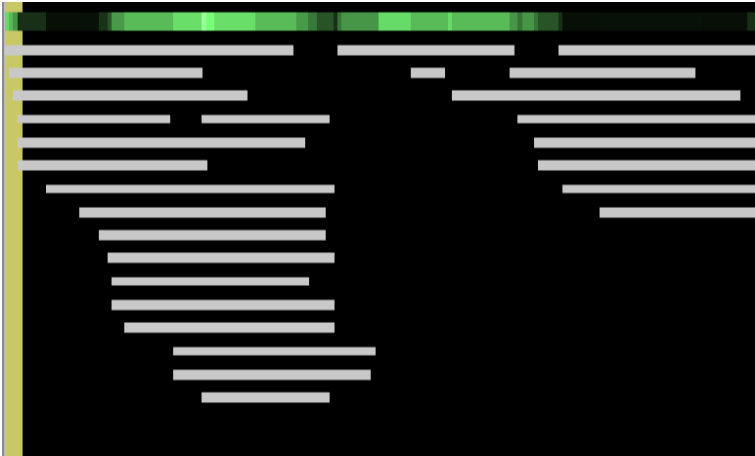
TVÉ01_E02+	ACAGAG-ATCGGTGGTCAGGAAA GTGGCA GCTCGAATAGCACCCBAGATCGGTCGTGAAGA
TVÉ02_E05+	ACCGAG-ATCGGTGGTGA
TVÉ01_H11+	ACCGAGCATCGGTGGTGAAGAAA GTG
TVÉ04_F07-	ACCGAA-ATCGGTGGTGAAGAAA GTGGCA GCTCGAATAGT
TVÉ03_F07-	ACAGAG-ATCGGTGGTGAAGAAA GTGGCA GCTCGAATAGCACCCBAGATCGGTCGTGAAGA
TVÉ10_A02+	ACCGAG-ATCGGTGGTGAAGAAA GTG
TVÉ05_D09+	ACCGAA-ATCGGTGGTGAAGAAA GTGGCA GCTCGAATAGCACCCBAGATCGGTCGTGAAGA
TVÉ05_E01+	ACCGAN-ATCGGTGGTGAAGAAA GTGGCA GCTCGA
TVÉ05_F04+	ACCGAG-ATCGGTGGTGAAGAAA GTGGCA GCTCGAATAGCACCCBAGATCGGTCGTGAAGA
TVÉ11_F02+	ACCGAG-ATCGGTGGTGAAGAAA GTGGCA GCTCGAATAGCAC
TVÉ03_D12-	ACAGAG-ATCGGTGGTGAAGAAA GTGGCA GCTCGAATAGCACCCBAGATCGGTCGTGAAGA
TVÉ10_F09+	ACCGAGATCGGTCGTGAAGA
TVÉ11_D02+	ACCGAGATCGGTCGTGAAGA
consensus	ACCGAG-ATCGGTGGTGAAGAAAGTGGCA GCTCGAATAGCACCCBAGATCGGTCGTGAAGA

Ensamble

- Ensamble Salmo Salar:

Numero de Lecturas	485834
Numero de Lecturas fwd	291492
Numero de Lecturas rev	194342
Mate pair	189426
Numero de Singlest	111051
Numero de Lecturas Ensamble	374781
Promedio Largo Lectura	650
Numero de Contig	49297
Promedio Largo Contig	1218
Desviación Estándar Largo Contig	480
Profundidad Promedio	7
Profundidad Máxima	726

Curación Ensamblas



Visualización Información

Salmo Salar Contig EST website - Mozilla Firefox

Archivo Editar Ver Historial Marcadores Herramientas Ayuda


http://bio-4.dim.uchile.cl/ssest/

Most Visited Smart Bookmarks Getting Started Latest Headlines TCB Group Software

Home | Navigating | Features | Links | Gene or mRNA search

Salmo Salar Contig EST website

Overview



Atlantic salmon, known scientifically as *Salmo salar*, is a species of fish in the family Salmonidae, which is found in the northern Atlantic Ocean and in rivers that flow into the Atlantic and the Pacific.

Distribution and habitat

Beginning around 1990 the rates of Atlantic salmon mortality at sea more than doubled, and by 2000 the numbers of Atlantic salmon had dropped to critically low levels. In the western Atlantic fewer than 100,000 of the important multi-sea-winter salmon were returning. Rivers of the coast of Maine, plus southern New Brunswick and much of mainland Nova Scotia saw runs drop precipitously, and even disappear. Beginning in the mid-1990s the Atlantic Salmon Federation in cooperation with partners were developing sonic tracking technology, and by 2008 the salmon have been tracked from rivers such as the Restigouche and the Miramichi as far along their migration routes as the Strait of Belle Isle, between Labrador and Newfoundland - and half way to feeding grounds in Greenland. For whatever reasons, possibly related to improvements in ocean feeding grounds, returns in 2008 have been very positive. On the Penobscot returns had been about 940 in 2007, and by mid-July 2008 the return was 1,938. Similar stories were played out in rivers from Newfoundland to Quebec. The problems at sea remain, and there is a concerted international effort called SALSEA, to find out more about the mortality at sea. It is organized by the North Atlantic Salmon Conservation Organization (NASCO). There are still residual problems from the past, including acid rain impacts in Nova Scotia, and dams in Maine, Quebec, New Brunswick, Newfoundland and throughout Europe, but the principal problem is the mortality levels at sea. Around the North Atlantic, efforts to restore salmon to their native habitats are underway and there is some slow but steady progress. Restoration and protection of the habitat itself is key to this process but issues of excessive harvest and competition with farmed and escaped salmon are also primary considerations. In the Great Lakes, Atlantic salmon have been introduced successfully, but the actual percentage of salmon reproducing naturally is very low. Most are stocked annually. Atlantic salmon were native to Lake Ontario but were extirpated by habitat loss and overfishing in the late 19th century. The state of New York has since been annually stocking its adjoining rivers and tributaries with the fish and in many cases does not allow active pursuit of the species. Wild salmon on entering rivers as adults have characteristically pointed fins which help scientists distinguish from farmed or escaped salmon.

- Salmo salar site version 0.0
- This sample site can be used to display research results and new annotations for Salmo Salar Contig EST database.

This website is under construction.

Tools

- [Blast Search](#) (BlastGraphic)
- [Contig Visualization](#) (GBrowse)
- [Comparative Browser](#) (CMap)
- [Genome Download](#)
- [Pathways Browser](#) (Salmo Salar Cyc)
- [Additional Tools...](#)

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This site powered by LBMG

Danio rerio

Danio rerio

version: 1.0

<u>Replicon</u>	<u>Total Genes</u>	<u>Protein Genes</u>	<u>RNA Genes</u>	<u>Pseudogenes</u>	<u>Size (bp)</u>
Chromosome 1 (90 contigs)	1211	1127	84	0	
Chromosome 2 (125 contigs)	1218	1105	113	0	
Chromosome 3 (111 contigs)	2276	1294	982	0	
Chromosome 4 (89 contigs)	2873	822	2051	0	
Chromosome 5 (136 contigs)	2087	1326	761	0	
Chromosome 6 (117 contigs)	1300	997	303	0	
Chromosome 7 (103 contigs)	1358	1239	119	0	
Chromosome 8 (121 contigs)	1780	1033	747	0	
Chromosome 9 (87 contigs)	979	843	136	0	
Chromosome 10 (71 contigs)	891	854	37	0	
Chromosome 11 (83 contigs)	861	807	54	0	
Chromosome 12 (113 contigs)	1176	835	341	0	
Chromosome 13 (102 contigs)	994	907	87	0	
Chromosome 14 (111 contigs)	2454	906	1548	0	
Chromosome 15 (101 contigs)	1251	934	317	0	
Chromosome 16 (107 contigs)	1483	1029	454	0	
Chromosome 17 (101 contigs)	1117	895	222	0	
Chromosome 18 (93 contigs)	893	795	98	0	
Chromosome 19 (86 contigs)	904	868	36	0	
Chromosome 20 (83 contigs)	1618	1076	542	0	
Chromosome 21 (111 contigs)	1936	900	1036	0	
Chromosome 22 (79 contigs)	1293	1037	256	0	
Chromosome 23 (90 contigs)	863	819	44	0	
Chromosome 24 (90 contigs)	983	648	335	0	
Chromosome 25 (83 contigs)	819	719	100	0	
Chromosome MT (1 contigs)	37	13	24	0	
Total:	34655	23828	10827	0	

Pathways:	123
Enzymatic Reactions:	837
Transport Reactions:	2
Polypeptides:	24229
Protein Complexes:	1
Enzymes:	913
Transporters:	6
Compounds:	702
Transcription Units:	0
tRNAs:	10211

Taxonomic lineage: cellular organisms, Eukaryota, Fungi/Metazoa group, Metazoa, Eumetazoa, Bilateria, Coelomata, Deuterostomia, Chordata, Craniata, Vertebrata, Gnathostomata, Teleostomi, Euteleostomi, Actinopterygii, Actinopteri, Neopterygii, Teleostei, Elopococephala, Clupeocephala, Otocephala, Ostariophysi, Ostariophysii, Cypriniformes, Cyprinidae, Rasbora, *Danio rerio*

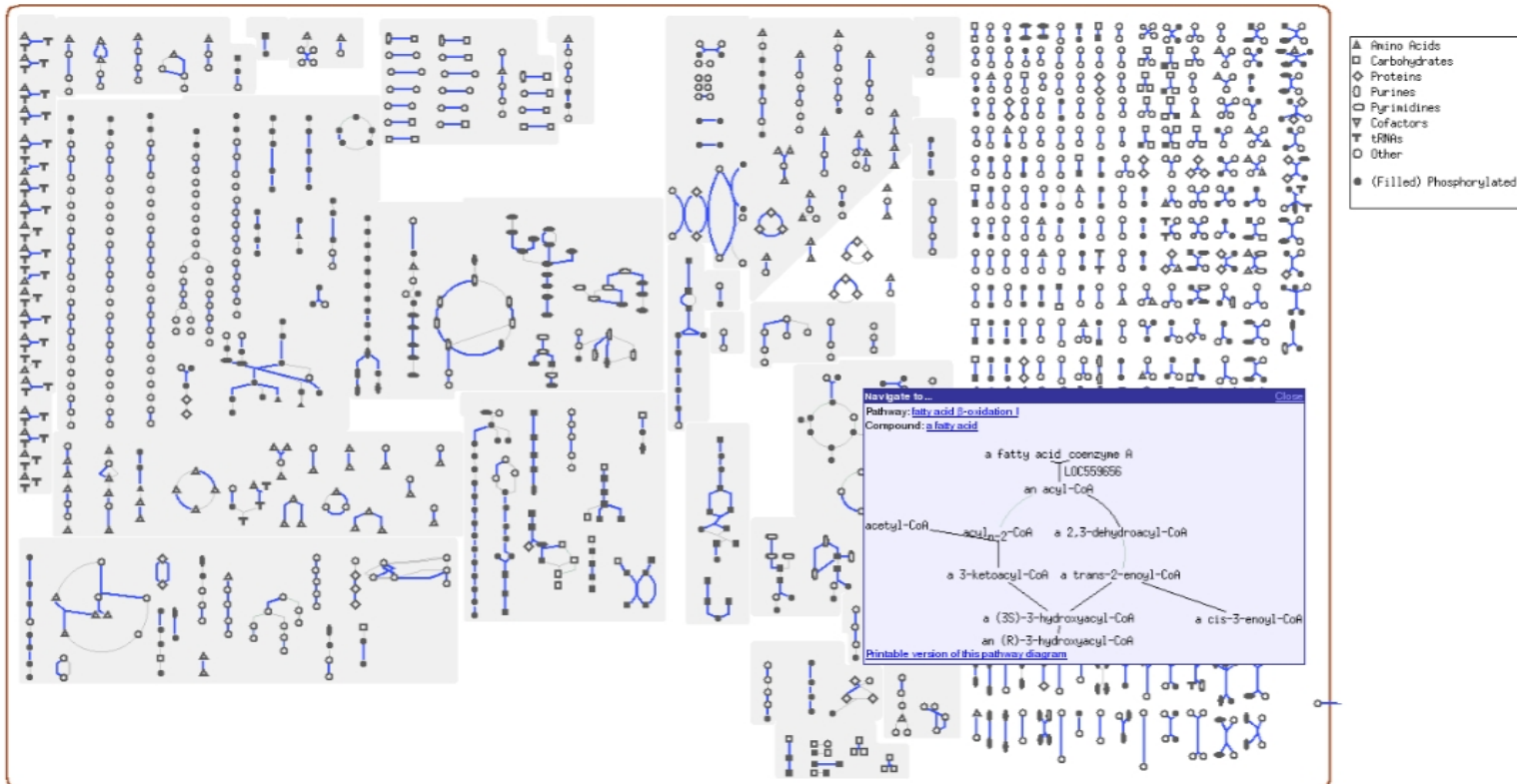
Unification Links: [NCBI-Taxonomy:7955](#)

Danio rerio

Overview of the *D. rerio* Metabolic Map

This diagram provides a schematic of all pathways of *D. rerio* metabolism in the daniozyc database. Nodes represent metabolites, with shape indicating class of metabolite (see key to right). Lines represent reactions. Move the mouse over a metabolite icon to identify it. Click on a metabolite icon to navigate to the metabolite page or a related pathway page.

- [Instructions](#)
- [Pathway Tools query page](#)
- [Omics Viewer: Paint omics data onto this diagram](#)
- [Species Comparison: Highlight reactions shared with other organisms](#)



Danio rerio

- Proteínas:

	Number	Percentage
hypothetical protein	7420	31,2
know function	16408	69,8
total proteins	23828	100